

www.trifas.org ISSN 1303-2712 Turkish Journal of Fisheries and Aquatic Sciences 16: 1025-1029 (2016) DOI: 10.4194/1303-2712-v16\_4\_29

#### SHORT PAPER

# A Preliminary Study on Protease Activity of Russian Sturgeon, Acipencer gueldenstaedtii Brandt and Ratzenburg, 1833, at Early Life Stages

# Kaya Gökçek<sup>1,\*</sup>, Tamás Szabó<sup>2</sup>, Cemal Alptekin<sup>3</sup>, Recep Kurt<sup>3</sup>, Yalçın Töre<sup>4</sup>, Béla Urbányi<sup>2</sup>

<sup>1</sup> Mustafa Kemal University, Agriculture Faculty Animal Sciences Department, Antakya, Hatay, Turkey.

<sup>2</sup> Szent Istvan University, Fish Culture Department, Gödöllö, Hungary.

<sup>3</sup> Iskenderun Technical University, Marine Sciences and Technology Faculty, Iskenderun, Hatay, Turkey.

<sup>4</sup> Şırnak University, Agriculture Faculty Animal Sciences Department, İdil, Şırnak, Turkey.

* Corresponding Author: Tel.: +90.534 4150716 ;	Received 12 May 2016
E-mail: kayagokcek@yahoo.com	Accepted 06 June 2016

#### Abstract

The fish feed industry continues to researches for optimum diet demands for candidate species culture. For this purpose, in vitro analyze methods may be more efficient than in vivo assays. This study includes two different stages; the first one is about the effect of commercial feeding protocol on protease enzyme alteration and the second one is about the inhibitory effects of different protein sources on early life proteases of Russian sturgeon, A. gueldenstaedtii, juveniles. In the first step, feeding with live prey and transition period to artificial feed significantly affected the daily amount of protease in digestive system (P<0.05). In the second part, some protein sources used in micro diets were tested in vitro for examination of their possible inhibitory effects on the proteases of Russian sturgeon larvae. The minimum inhibitory effect was obtained from fish meal (15.44%), but however, soybean protein concentration, soybean meal, corn gluten and rice bran inhibited the proteases significantly higher than FM and its combinations (63.55, 71.81, 72.24, and 80.77%, respectively). Additionally, dual combinations between fish meal and soybean meal/soybean protein concentration with the ration of three to one (3:1) was moderate (26.38 and 22.13 %), whereas blood meal extremely produced a 97.28% inhibitory ratio.

Keywords: Russian sturgeon, protease activity, inhibition, feed ingredients.

Rus Mersin Balığının, Acipencer gueldenstaedtii Brandt ve Ratzenburg, 1833, Yaşamlarının İlk Dönemlerindeki Proteaz Aktivitesi Üzerine Bir Ön Çalışma

#### Özet

Balık yemi endüstrisi, aday türlerin optimum rasyon ihtiyacını belirlemek için araştırmalarına devam etmektedir. Bu amaçla yapılan in vitro analiz metotları, in vivo metotlara göre daha verimli olabilir. Bu çalışma iki farklı çalışmadan oluşmaktadır; bunlardan birincisi ticari besleme protokolünün proteaz enzimlerinin değişimi üzerine olan etkisini, ikinci çalışma ise farklı protein kaynaklarının Rus mersini, A. gueldenstaedtii, jüvenillerinin erken dönem proteaz aktivitiesi üzerine olan inhibisyon etkisini belirlemeyi amaçlamıştır. İlk çalışmada, canlı yem ile besleme ve karma yeme geçiş periyodunun, sindirim sistemindeki günlük proteaz miktarını istatistiki olarak etkilediği tespit edilmiştir (P<0.05). İkinci çalışmada ise, mikro yemlerde kullanılan bazı protein kaynakları, Rus mersini jüvenillerinin proteaz enzimlerini olası inhibe edici etkilerini belirlemek için in vitro olarak test edilmiştir. En düşük inhibisyon etkisi balık ununda (%15.44) gözlemlenmiş, ancak buna karşın soya protein konsatresi, soya unu, mısır glüten ve pirinç kepeği proteaz enzimlerini balık unu ve diğer hammaddelerle yaptığı kombinasyonlardan istatistiki açıdan daha yüksek oranda inhibe etmiştir (sırasıyla % 63.55, 71.81, 72.24 ve 80.77). Buna ek olarak, balık ununun soya unu/soya protein konsantresi ile 1:3 oranındaki kombinasyonları kabul edilebilir seviyede inhibe edici bulunurken (%26.38 ve 22.13), kan ununun aşırı seviyede inhibe edici olduğu tespit edilmiştir (%97.28).

Anahtar Kelimeler: Rus mersini, proteaz aktivitesi, inhibisyon, yem hammaddeleri.

## Introduction

Sturgeons are one of the oldest living vertebrates and generally called "living fossils". Populations are found primarily in cold and temperate regions of the Northern hemisphere. All sturgeon species are endangered due to overfishing, river damming, regulation, deepening and strengthening of river banks (Napora-Rutkowski et al., 2009). Interest in sturgeon species mainly focused on caviar, smoked fish, soup, isinglass (collagen), ivory and leather.

Although endogenous production of digestive enzymes by larvae is emphasized to be adequate for exogenous live or artificial feeds (Hofer and Köck, 1989; Kolkovski, 2001), the selection of the most suitable feed ingredients for microfeeds and

<sup>©</sup> Published by Central Fisheries Research Institute (CFRI) Trabzon, Turkey in cooperation with Japan International Cooperation Agency (JICA), Japan

manufacturing are very important for candidate specie's culture. Recently, the plant protein sources were intensively started to be used in commercial fish feeds instead of fishmeal to protect natural fish populations which are exploited for fishmeal production. On the other side, fish farmers complain for high feed conversion ratios and low growth performance. It is thought that this situation is probably caused by plant protein sources due to having inadequate amino acid profiles (Halver and Hardy, 2002).

Aquaculture research results concerning breeding, larval rearing and digestive enzymes were already presented for some Acipencer species (Willot et al., 2001; Gawlicka et al., 2002; Furne et al., 2005; Napora-Rutkowski et al., 2009; Babaei et al., 2011; Sanz et al., 2011; Noori et al., 2012) and commercial feeds are already produced by some companies. However, the effect of commercial feeding protocol on enzyme activity, inhibitory effects and bioavailability of combination of different protein sources on protease activity of Russian sturgeon is still obscure. In the present work, in vitro methods were examined to assess the variance of protease levels according to feed types and the interaction between proteases and different protein sources that can be used in preparation of microfeeds for Russian sturgeon.

#### **Materials and Methods**

#### Source of Fry

Fertilized sturgeon eggs were imported from Russia and hatched in commercial fish farm in Homokmègy, Hungary (Rideg & Rideg Ltd.). After yolk sac absorption, larvae were transferred and stocked into recirculation system. The water temperature was  $17\pm1^{\circ}$ C during experiment. Commercial feeding protocol (Figure 1) was applied for 30 days and 30 days-old larvae were sampled into 3 ml eppendorf tubes. Samples were rinsed in distilled water and stored at  $-80^{\circ}$ C until protease and inhibition analyses. The whole body of larvae were homogenized to get enzyme extracts by centrifugation (12000 g, 20 min,  $4^{\circ}$ C).

#### **Determination of Protease Activities**

Casein (10 mg ml<sup>-1</sup>) was used as substrate and dissolved in 50 mM Tris-HCl buffer at pH 9 (Walter, 1984). The substrate and the enzyme extracts of larvae were incubated at 37°C for 1 hour. Then, 500  $\mu$ l trichloroacetic acid (TCA) (120 g L<sup>-1</sup>) was used to stop the reaction. The mixture was incubated at 4°C for 10 minutes and after centrifugation (8000 g, 15 min, 4°C) the absorbance was recorded at 280 nm. One unit of enzyme activity was defined as 1  $\mu$ g of tyrosine released per minute (Hazman and Gökçek, 2014). The soluble protein concentrations of larvae were determined according to Bradford (1976).

#### Determination of Inhibition Effects of Protein Sources on Proteolitic Activity

A modified method of Garcia-Carreno (1996) was used to determine inhibition effects of fish meal (FM), soybean meal (SM), soybean protein concentration (SPC), corn gluten (CG), rice bran (RB), blood meal (BM) and combination of FM/SM (1:3, 1:1, 3:1), FM/SPC (1:3, 1:1, 3:1) and SM/MG (1:3, 1:1, 3:1) as protein sources on protease activity of 30 day-old juvenile. Different feed ingredients and protease enzyme extracts were pre-incubated and the residual activity was measured. The extracts (20 µL) were incubated with protein solutions (20 µL) in HCl Tris buffer (500 µL) for 60 min at 25°C (pH 9.0). In the control group, same amount of distilled water was used instead of protein solutions in the mixture. Afterwards, 100 µL casein was added and the mixture was incubated for 120 min at room temperature. Finally, 500 µL TCA (120 gL-1) was added to stop reaction and after centrifugation (8000 g, 15 min, 4°C) the absorbance was recorded at 280 nm. TCA was added prior to casein in blanks (Alarcon et al., 1999).

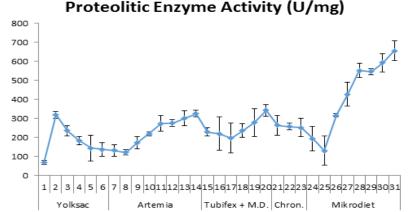


Figure 1. Feeding protocol and daily chances of digestive protease activity on Russian sturgeon at first month.

#### Statistical Methods

Kolmogorov-Smirnov normality test and Bartlett's test for homogeneity were used prior to comparison of data. A one-way ANOVA test was applied for comparisons by using SPSS 15.0 (Hazman and Gökçek, 2014).

#### Results

The feeding protocol and daily chances of digestive protease activity on Russian sturgeon prepost larvae and juveniles were presented in Figure 1. By the possible increasing of cytosolic enzyme secretion on 2nd day after hatching (DAH), the protease activity reached to the highest level in the yolk sac stage. Then, the enzyme activity decreased to the minimum level by decreasing of the total mass of yolk sac. Exogenous feeding with Artemia directly affected the enzyme activity and it increased till the DAH 14. After this point, post larvae were fed with tubifex+M.D. mixture, however larvae unwillingly accepted this new feed type between DAH 15th and 18th. Although larvae were corresponded to new feed habit, the same situation was observed on the feeding period with Chironomid+M.D. between DAH 21st and 23rd. The second lowest enzyme secretion was obtained on the DAH 25. By the development of digestive tract, microdiet feed was accepted by juveniles and the total protease activity started to increase after DAH 26th.

The protease activity was measured  $592.74\pm47.61$  U/mg on 30 days-old fry. The inhibitory degree of feed ingredients on protease activity was given in Table 1. The lowest inhibition was observed from FM ( $15.44\pm5.59\%$ ) and inhibition produced by other protein sources were relatively high. Combination of two different feed ingredients inhibited relatively higher such as  $26.38\pm2.02$  and  $22.13\pm2.69\%$  from FM/SM (3:1) and FM/SPC (3:1),

respectively. The highest inhibition of juvenile protease activity was induced by BM (97.28±2.31).

## Discussion

In this study, the mouth of Russian sturgeon larvae was opened on 9th day and the protease activity was increased by the presence of Artemia in the culture medium. Sanz et al. (2011) stated that although the acidic protease activity were detected at the beginning of the exogenous feeding, the increase of enzyme activity should be exogenous originated because gastric glands didn't have the granules of zymogen secretion in A. naccarii yet. Controversially, it has been suggested by several researchers that exogenous enzymes are unnecessary for digestion in the early larval stage (Segner et al., 1993; Kim et al., 2001) but the live food may trigger the secretion of proteases (Zambanino Infante and Cahu, 2001). Afterwards, feeding firstly with tubifex+ M.D. and Chironomids+M.D. negatively affected the digestion and absorption capabilities due to possibly inadequate enzyme secretion (Faulk et al., 2007). Then, the protease activity was on the rise in an orderly manner after DAH 25. This is because the juvenile developed anatomically similar digestive system as like adults (Sanz et al., 2011).

The present study demonstrated that BM significantly reduced (97.28%) the activity of digestive proteases in 30 day-old Russian sturgeon. On the other, FM showed the lowest (15.44%) inhibitory effect on protease activity. Alptekin (2016) stated the same results for FM (15.34%) in Siberian sturgeon, A. baerii. In their study, there were no statistical differences between FM and SPC (P<0.05), even though the SPC showed a lower inhibitory effect on protease activity (14.45%). In the present study, all other single ingredients negatively affected the digestive activity in contrast to FM. Similar results were obtained when commercial ingredients were

Table 1.	Protein	sources a	and	inhibition	degree of	protease activity	v

Protein Sources	Inhibition degree of protease activity ( $\pm$ SD)
Fish Meal (FM)(69% CP)	15.44±5.59ª
Soybean Meal (SM)(48% CP)	71.81±3.82 <sup>e</sup>
Soybean Protein Concentration (SPC) (58% CP)	63.55±5.46 <sup>de</sup>
Corn Gluten (CG)(60% CP)	72.24±4.17 <sup>e</sup>
Rice Bran (RB)(n/a)	$80.77{\pm}2.28^{ m f}$
Blood Meal (BM)(75.5% CP)	$97.28 \pm 2.31^{g}$
FM/SM (1:3)	$76.81 \pm 4.39^{\text{ef}}$
FM/SM (1:1)	$59.24 \pm 8.45^{d}$
FM/SM (3:1)	$26.38{\pm}2.02^{ m b}$
FM/SPC (1:3)	$62.46 \pm 6.35^{d}$
FM/SPC (1:1)	35.75±5.67°
FM/SPC (3:1)	$22.13 \pm 2.69^{b}$
SM/CG (1:3)	$69.02 \pm 6.92^{d}$
SM/CG (1:1)	$70.85 \pm 9.89^{de}$
SM/CG (3:1)	$76.95 \pm 4.52^{ m ef}$

n/a: not available

Means with the same superscripts are not significantly different (P<0.05)

tested for northern pike juveniles (Töre *et al.*, 2014). Whereas, the combination of FM/SM (3:1) and FM/SPC (3:1) showed acceptable results (26.38 and 22.13 %, respectively). By increasing of FM ration in both mixture, the inhibition effect clearly reduced probably due to the changing of amino acid composition.

Micro feeds for Sturgeons are already commercially produced by several feed companies. Although these feeds are accepted by fish, sometimes poor growing or nutritional originated health problems can be occurred. This is probably due to undeveloped digestive tract in early stages of larvae or juveniles. On the other hand, some researchers claimed that some specie' digestive enzyme secretion, especially a high protease and amylase activity, were already existed since early stage of development (Cahu and Zambonino, 1995; Moyano et al., 1996) and it has been hypothesized that this is due to inhibition of enzymes by inadequate protein sources (Alarcon et al., 1999). Also, limited intake of amino acids in charged to produce new body proteins may be caused the presence of enzyme inhibitors in artificial feeds (Garcia-Carreno, 1996). In this study, no reduction were obtained, even though dry air were used to extinguish possible inhibitory. By this kind of in vitro assays, the protein sources in used for adult fish (Grabner 1985; Dimes et al., 1994) and shrimp (Lana and Pan, 1993; Ezquerra, 1997) were already evaluated to prove that these ingredients are useful or not, however this is the first evaluation for a sturgeon specie.

In conclusion, according to analyze results, feeding larvae with Tubifex and Chronomids without M.D. supplementation before DAH 25 can give better results for adequate secretion of proteases. Also, it is clear that the plant protein sources were negatively affected protease activity on DAH 30. The information from in vitro assays may preliminary provide an explanation for why plant protein sources gave poor results for Russian sturgeons. However, to get a better knowledge about the effect of different feed ingredients, the determination of the degree of hydrolysis and sequential analysis of protein hydrolysis products should be examined in the future.

#### References

- Alarcon, F. J., Moyano, F. J., Diaz, M., Fernandez-Diaz, C., Yufera, M., 1999. Optimization of the protein fraction of microcapsules used in feeding of marine fish larvae using *in vitro* digestibility techniques. Aquaculture Nutrition, 5: 107-113. doi: 10.1046/j.1365-2095.1999.00093.x
- Alptekin, C., Gökçek, K., 2016. Balık Yemlerinde Kullanılan Farklı Protein Kaynaklarının Sibirya Mersini (Acipencer baerii Brandt 1869) Jüvenillerinin Proteaz Aktivitesi Üzerine Etkileri. Yunus Araştırma Bülteni, doi:10.17693/yunus.32866
- Babaei, S. S., Kenari, A. A., Nazari, R., Gisbert, E., 2011. Developmental changes of digestive enzymes in

Persian Sturgeon (*Acipencer percius*) during larval ontogeny. Aquaculture, 318: 138-144. doi:10.1016/j.aquaculture.2011.04.032

- Bradford, M. M. 1976. A rapid sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dry binding. Anal. Biochem., 72, 248-254.
- Cahu, C. L. and Zambonino, J. L., 1995. Maturation of the pancreatic and intestinal digestive functions in sea bass (*Dicentrarchus labrax*): effect of weaning with different protein sources. Fish Physiol. Biochem., 14: 431-437.
- Cho, C.Y. 1987. La energia en la nutricion de los peces. In: Nutricion en Acuicultura, vol. II (ed. By de la Monteros Espinosa nad Lanarta U.), pp. 197-244. CAICYT, Madrid, Spain.
- Dimes, L. E., Haard, N. F., Dong, F. M., Rasco, B. A., Forster, J. P., Fairgrieve, W. T., Arndt, R., Hardy, R. W., Barrows, F. T., Higs, D. A., 1994. Estimation of protein digestibility. II. *In vitro* assay of protein in salmonid feeds. Comp. Biochem. Physiol. 108A: 363-370. doi:10.1016/0300-9629(94)90107-4
- Ezquerra, J. M., 1997. Enzyme technology in aquaculture: *in vitro* evaluation of enzymatic digestibility of dietary proteins by pH-stat for the optimization of diets for cultured *Penaeus vannamei*. Ph.D Thesis, CIBNOR, La Paz, Mexico.
- Faulk, C. K., Benninghoff, A. D., Holt, G. J., 2007. Ontogeny of the gastrointestinal tract and selected digestive enzyme in cobia *Rachycentron canadum* (L.). J. Fish Biol. 70:567-583. doi: 10.1111/j.1095-8649.2007.01330.x
- Furne, M., Hidalgo, M. C., Lopez, A., Garcia- Gallego, M., Morales, A. E., Domezain, A., Domezaine, J., Sanz, A., 2005. Digestive enzyme activities in Adriatic sturgeon Acipencer naccarii and rainbow trout Oncorhynchus mykiss. A comperative study. Aquaculture, 250: 391-398. doi:10.1016/j.aquaculture.2005.05.017
- Garcia-Carreno, F. L., 1996. Proteinase inhibitors. Trends Food Sci. Technol., 7: 197-204. doi:10.1016/0924-2244(96)10023-6
- Gawlicka, A., Herold, M. A., Barrows, F. T., de le Noue, J., Hung, S. S. O., 2002. Effect of dieatary lipids on growth, fatty acid composition, intestinal absorbtion and hepatic storage in white sturgeon (*Acipencer transmontanus*) larvae. J. Appl. Icthyol., 18: 673-681. doi: 10.1046/j.1439-0426.2002.00371.x
- Grabner, M., 1985. An in vitro method for measuring protein digestibility of fish feed components. Aquaculture, 48: 97-110. doi:10.1016/0044-8486(85)90097-3
- Hazman, B., Gökçek, K., 2014. The Effect of Different First Feeds on Proteolitic Activity of the Northern Pike, *Esox lucius* Linneaus 1758, Post-Larvae. *Turkish Journal of Fisheries and Aquatic Sciences* 14 (4): 875-878. doi: 10.4194/1303-2712-v14\_4\_04
- Halver, J.E., Hardy, R.W., 2002. Fish Nutrition. Academic Press, USA. pp. 417-427.
- Hofer, K., Köck, G. 1989. Method for quantitative determination of digestive enzymes in fish larvae. Polish Archives of Hidrobiology, 36, 439-441.
- Kim, B. G., Divakaran, S., Brown, C. L., Ostrowski, A. C., 2001. Comparative digestive enzyme ontogeny in two larval fishes: Pacific threadfin (*Polydatilus sexfilis*) and bluefin trevally (*Caranx melampygus*). Fish Physiol. Biochem. 24: 225-241.

- Kolkovski, S., 2001. Digestive enzymes in fish larvae and juveniles- implications and applications to formulated diets. Aquaculture 200: 181-201.
- Lan, C. C. and Pan, B. S., 1993. In vitro digestibility sitimulating the proteolysis of feed protein in the midgut gland of grass shrimp *Penaeus monodon*. Aquaculture, 109: 59-70. doi:10.1016/0044-8486(93)90486-I
- Moyao, F. J., Diaz, M., Alarcon, F. J., Saraquete, M. C., 1996. Characterization of digestive enzyme activity during larval development of gilthead seabream (*Sparus auraus*). Fish Pyhsiol. Biochem., 15: 121-130.
- Napora- Rutkowski, L., Kamaszewski, M., Bielawski, W., Ostaszewska, T., Wegner, A., 2009. Effects of starter diets on pancreatic enzyme activity in juvenile starlet (*Acipencer ruthenus*). The. Isr. Jour. Aqua.-Bamidgeh, 61 (2): 143-150.
- Noori, F., Gilbert, V. S., Sorgeloos, P. 2012. Preliminary study on the activity of protease enzymes in Persian sturgeon (*Acipencer persicus* Borodin, 1897) larvae in response to different diets: effects on growth and survival. Aquaculture Research, 43, 198-207. doi: 10.1111/j.1365-2109.2011.02816.x
- Sanz, A., Llorente, J. L., Furne, M., Ostos-Garrido, V., Carmona, R., Domezain, A., Hidalgo, M. C., 2011. Digestive enzymes during ontogeny of the sturgeon

Acipencer naccarii: intestine and pancreas development. J. Appl. Ichthyol., 27: 1139-1146. doi: 10.1111/j.1439-0426.2011.01864.x

- Segner, H., Rösch, R., Verreth, J., Win, U., 1993. Larval nutrition physiology: studies with *Clarias gariepinus*, *Coregonus lavaretus* and *Scophthalmus maximus*. J. World Aqua. Soc. 24: 121-134. doi: 10.1111/j.1749-7345.1993.tb00001.x
- Töre, Y., Gökçek, K., Alptekin, C., Szabo, T. ve Urbanyi, B., 2014. Turna, *Esox lucius* Linneaus 1758, Balığı Larvalarının Proteaz Aktivitesi Üzerine Ticari Yem Hammaddelerinin İnhibisyon Etkilerinin Tespiti. Doğu Anadolu 5. Su Ürünleri Sempozyumu. 31 Mayıs-2 Haziran 2014, Elazığ.
- Walter, H.E. 1984. Proteinases: methods with hemoglobin, casein and azocoll as sub-strates, In: Methods of Enzymatic Analysis (Bergmeyer, H.J. ed.), 5, 270-277.
- Willot, P., Sabeau, L., Gessner, J., Arlati, G., Bronzi, P., Gulyas, T., Berni, P., 2001. Sturgeon farming in Western Europe: recent developments and perspectives. Aquat. Living Resourc., 14: 367-374. doi:10.1016/S0990-7440(01)01136-6
- Zambonino Infante, J.L., Cahu, C.L. 2001. Ontogeny of the gastrointestinal tract of marine fish larvae. Comp. Biochem. Physiol. C, 130: 477-487. doi:10.1016/S1532-0456(01)00274-5