

# The Changes in Biochemical Compositions of Five Different Macroalgae and Seagrass (Halophila stipulacea (Forsskal) Ascherson 1867) Collected from Iskenderun Bay

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Article HistorReceived:14.03.2022Accepted:31.08.2022Published:15.12.2022Research Article	Abstract –In present study, biochemical compositions (ash, lipid and protein) of five different macroalgae ((Green Macroalgae-GMA ( <i>Chaetomorpha linum</i> and <i>Caulerpa prolifera</i> ), Red Macroalgae-RMA ( <i>Pterocladiella capilacea</i> ), Brown Macroalgae-BMA ( <i>Sargassum vulgare</i> and <i>Ericaria amentacea</i> )) and Angiosperm/Seagrass ( <i>Halophila stipulacea</i> ) collected from Iskenderun Bay were investigated. The differences observed between biochemical compositions such as ash, lipid and protein of five macroalgae species and Angiosperm/Seagrass ( <i>Halophila stipulacea</i> ) were statistically significant (p<0.05). The lowest and highest ash, lipid and protein values of five macroalgae were 12.19±1.15% ( <i>Caulerpa prolifera</i> )- 21.38±1.53% ( <i>Ericaria amentacea</i> ), 1.74±0.19% ( <i>Caulerpa prolifera</i> )- 5.83±0.68% ( <i>Ericaria amentacea</i> ), 5.56±0.06% ( <i>Chaetomorpha linum</i> )- 11.45±0.53% ( <i>Sargassum vulgare</i> ), respectively. Ash, lipid and protein values of Angiosperms/Seagrass ( <i>Halophila stipulacea</i> ) were determined as 14.56±2.08%, 3.16±0.48% and 8.11±0.07%, respectively. Protein value of Angiosperms/Seagrass ( <i>Halophila stipulacea</i> ) was higher than those of (GMA ( <i>Chaetomorpha linum</i> and <i>Caulerpa prolifera</i> ) but not RMA( <i>Pterocladiella capillacea</i> ) and BMA ( <i>Sargassum vulgare</i> and <i>Ericaria amentacea</i> ). Lipid value of Angiosperms/Seagrass ( <i>Halophila stipulacea</i> ) are important for the evaluation of potential sources for commercial and human consumption. In addition, biochemical compositions of tested macroalgae and seagrass could make important contributions to feed formulations and functional foods in future.
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Keywords-Green algae, brown algae, red algae, seagrass, biochemical compositions

## **1. Introduction**

Macroalgae known as primary producers are important for the ocean ecosystem. They include nutritional compounds used different industries such as food, agriculture, feed, cosmetic, medicine, pharmacy. Ak (2015)

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showed that macroalgae have contain different photosynthetic pigments such as Chlorophyta (Green Macroalgae-GMA), Rhodophyta (Red Macroalgae-RMA), Ochrophyta (Brown Macroalgae-BMA). Researchers revealed that biological activity potentials, fatty acids, protein and mineral substances of macroalgae were high levels (Dawczynski, Schubert & Jahreis 2007; Wells et al., 2017). Burtin (2003) and Benjama & Masniyom (2011) reported that GMA and RMA have higher protein levels (10–30%) than BMA species (5–15%). Dawczynski et al. (2007) showed that proximate compositions of the mentioned GMA, RMA and BMA change according to the geographical distribution, species, season, water temperature, salinity, light and nutrients and mineral availability.

Caulerpa is GMA genera belonging to Caulerpaceae family. Invasive algae have important effects on the ecosystems. *Caulerpa taxifolia* is an invasive alga found in the Mediterranean Sea.

Seagrasses provide an important habitat for fish and crustaceans as well as ecological contribution (Fourqurean et al., 2012). It is known that, seagrasses represent to two families, Potamogetonaceae and Hydrocharitaceae, (Hemminga & Duarte, 2000). *Halophila stipulacea* is seagrass species belonging to the family Hydrocharitaceae. There are 10 species of the genus Halophila. *Halophila stipulacea* was first observed in the Mediterranean in 1894 (Lipkin, 1975). Due to its invasive nature, there is increasing interest in understanding the ability to tolerate various environmental conditions. (Malm, 2006).

Biochemical compositions (ash, lipid, protein) of macroalgae and seagrasses are important to determine for industrial potentials. There is need to knowledge about analysis of proximate compositions of macroalgae and seagrasses belonging to different regions. Studies about nutritional composition macroalgae and seagrass tested in this study is not adequate in Iskenderun Bay. Therefore, we aimed to investigate the biochemical compositions (ash, lipid and protein) of five different macroalgae ((GMA (*Chaetomorpha linum* and *Caulerpa prolifera*), RMA(*Pterocladiella capillacea*), BMA (*Sargassum vulgare* and *Ericaria amentacea*) and Angiosperms/Seagrass (*Halophila stipulacea*) gathered from Iskenderun Bay.

## 2. Materials and Methods

Macroalgae and Seagrass were gathered from the coasts of İskenderun Bay (Kale and Arsuz regions in July 2019). Five macroalgae species ((GMA (*Chaetomorpha linum (O.F.Müller*) Kützing 1845 and *Caulerpa prolifera (Forsskal) J.V.Lamouroux 1809*), RMA(*Pterocladiella capillacea (S.G.Gmelin) Santelices & Hommersand 1997*), BMA (*Sargassum vulgare C.Agardh, 1820* and *Ericaria amentacea (C.Agardh) Molinari & Guiry 2020*)) and Angiosperms/Seagrass (*Halophila stipulacea (Forsskal) Ascherson 1867*) were identified. The identification studies of macroalgae and seagrass were carried out with Olympus brand SZX16 model stereo zoom and BX51 model binocular light microscopes (Taşkın & Öztürk, 2013; Rodríguez-Prieto, Ballesteros, Boisset & Afonso-Carrillo, 2013; Cormaci, Furnari & Alongi, 2014)

## 2.1. Preparation of the samples

Five macroalgae and Angiosperms/Seagrass (*Halophila stipulacea*) collected from the coasts of Iskenderun Bay were rinsed in distilled water and then drained. Thus, sand and other unwanted objects were removed. The next step was to dry the samples at  $60^{\circ}$ C for 3 hours using a laboratory oven (Pradana, Prabowo, Hastuti, Djaeni & Prasetyaningrum, 2019). Thoroughly dried samples were ground with a laboratory type mixer and then, stored at  $-20^{\circ}$ C until analyses.

## 2.2. Biochemical compositions

Biochemical analyses of five macroalgae species ((GMA (*Chaetomorpha linum* and *Caulerpa prolifera*), RMA(*Pterocladiella capillacea*), BMA (*Sargassum vulgare* and *Ericaria amentacea*)) and

Angiosperms/Seagrass (*Halophila stipulacea*) gathered from the coasts of Iskenderun Bay were made according to the AOAC (2005) and Bligh & Dyer (1959) procedures.

#### 2.3. Statistical analysis

Ash, lipid and protein results of five macroalgae species ((GMA (*Chaetomorpha linum* and *Caulerpa prolifera*), RMA(*Pterocladiella capillacea*), BMA (*Sargassum vulgare* and *Ericaria amentacea*)) and Angiosperms/Seagrass (*Halophila stipulacea*) were submitted as mean  $\pm$  standard error (SE). Statistical comparisons were were made by OneWay Analysis (ANOVA) using SPSS 12. Differences were considered statistically significant when p<0.05.

#### 3. Results and Discussion

The purpose of study was to reveal biochemical compositions (ash, lipid and protein) of five different macroalgae ((GMA (*Chaetomorpha linum* and *Caulerpa prolifera*), RMA(*Pterocladiella capillacea*), BMA (*Sargassum vulgare* and *Ericaria amentacea*)) and Angiosperms/Seagrass (*Halophila stipulacea*). The biochemical compositions of five macroalgae and Angiosperms/Seagrass (*Halophila stipulacea*) are summarized in Table 1.

#### Table 1

<b>Biochemical</b>	compositions of	f macroalgae ai	nd seagrass (	(mean+SE)
Differentiation	compositions o	i macioaigae a	nu scagrass	mean±5L)

	Proximate Compositions			
GMA*	Ash (%)	Lipid (%)	Protein (%)	
Chaetomorpha linum	$17,68 \pm 0,33^{bc}$	4,84±1,68 <sup>b</sup>	5,56±0,06ª	
Caulerpa prolifera	12,19±1,15ª	$1,74{\pm}0,19^{a}$	6,70±0,07 <sup>b</sup>	
RMA**				
Pterocladiella capillacea	20,31±0,63°	3,11±0,53 <sup>ab</sup>	9,62±0,35 <sup>d</sup>	
BMA***				
Sargassum vulgare	$13,19 \pm 0,15^{a}$	4,31±0,42 <sup>ab</sup>	11,45±0,53e	
Ericaria amentacea	21,38±1,53°	5,83±0,68 <sup>b</sup>	9,75±0,07 <sup>d</sup>	
Angiosperms/Seagrass	Ash (%)	Lipid (%)	Protein (%)	
Halophila stipulacea	14,56±2,08 <sup>ab</sup>	$3,16\pm0,48^{ab}$	8,11±0,07°	

Different letters between the columns indicate significant difference at 5% by Duncan multiple range test.

\* Green Macroalgae; \*\* Red Macroalgae; \*\*\* Brown Macroalgae

The values found between biochemical compositions such as ash, lipid and protein of five macroalgae species and Angiosperms/Seagrass (Halophila stipulacea) were statistically significant (p<0.05). The lowest and highest ash, lipid and protein values of five macroalgae were 12.19±1.15% (Caulerpa prolifera)-21.38±1.53% (Ericaria amentacea), 1.74±0.19% (Caulerpa prolifera)- 5.83±0.68% (Ericaria amentacea), 5.56±0.06% (Chaetomorpha linum)- 11.45±0.53% (Sargassum vulgare), respectively. Ash, lipid and protein values of Angiosperms/Seagrass (Halophila stipulacea) were determined as 14.56 ±2.08%, 3.16±0.48% and 8.11±0.07%, respectively. Ash, lipid and protein values of *Ericaria amentacea* were the highest except for protein value of BMA Sargassum vulgare. Protein values of BMA were higher than those of GMA (Chaetomorpha linum and Caulerpa prolifera), RMA(Pterocladiella capillacea) and Angiosperms/Seagrass (Halophila stipulacea). Protein value of Angiosperms/Seagrass (Halophila stipulacea) was higher than those of GMA (Chaetomorpha linum and Caulerpa prolifera) but not RMA (Pterocladiella capillacea) and BMA (Sargassum vulgare and Ericaria amentacea)). Lipid value of Angiosperms/Seagrass (Halophila stipulacea) was similar to RMA (Pterocladiella capillacea). Ash and lipid values of Caulerpa prolifera except for protein value were lower than that of *Chaetomorpha linum*. Ash and lipid values of *Ericaria* amentacea except for protein value were lower than that of Sargassum vulgare. Ash and protein values of RMA (Pterocladiella capillacea) except for lipid value were higher than those of GMA (Chaetomorpha linum and Caulerpa prolifera) and Angiosperms/Seagrass (Halophila stipulacea).

According to the results of our study, protein levels of RMA(*Pterocladiella capillacea*), BMA (*Sargassum vulgare* and *Ericaria amentacea*)) were higher than those of GMA (*Chaetomorpha linum* and *Caulerpa pro-lifera*). Protein value of RMA(*Pterocladiella capillacea*) were similar to *Ericaria amentacea* but lower than that of BA (*Sargassum vulgare*).

It is known that, GMA and RMA contain higher protein contents (10–30%) than BMA (5–15%) (Burtin, 2003; Benjama & Masniyom, 2011). Dawczynski et al. (2007) revealed that protein levels of RMA was higher than those of BMA. However, Aras & Sayın (2020) showed that the protein ratio of *Ellisolandia elongata*, which is the only RMA species is similar to *Sargassum vulgare* and lower than *Dictyota dichotoma* from BMA.

Wahbeh (1997) showed that the protein content of the macroalgae *Padina pavonica* collected at the beach in Aqaba, Jordan was 17.4%. Tabarsa et al. (2012) found a protein content of 11.83% in *Padina pavonica*, which they collected in April in southern Iran (Persian Gulf). Ozgun & Turan (2015) showed that the protein levels of eight brown macroalgae collected from Iskenderun Bay was varied from 2.897±0.373% to 6.519±0.432%. Gür (2015) determined that protein value of *Dictyota dichotoma* was between 4.42-6.15%. Pakawan, Suriyan, Kriengkrai, & Jintana, (2015) biochemical compositions of 9 macroalgae species belonging to the GMA and RMA *Chaetomorpha crassa, Chaetomorpha linum, Ulva rigida, Caulerpa racemosa, Caulerpa brachypus, Caulerpa lentillifera, Caulerpa taxifolia, Gracilaria tenuistipitata and Gracilaria fisheri* showed as 12.68–33.83% protein. Uslu et al. (2021) determined that the protein amounts of the macroalgae *Sargassum vulgare* and *Cystoseira compressa* were 6.29±0.12% and 9.50±0.3%, respectively. Protein levels of *Sargassum vulgare* and *Cystoseira compressa* revealed by Uslu et al. (2021) were lower and similar to report from current study, respectively.

Protein levels of BMA (*Sargassum vulgare* and *Ericaria amentacea*)) tested in current study was lower than those of protein levels revealed by Wahbeh (1997) and similar to Tabarsa et al. (2012). However, protein levels of BMA tested were higher than those of Ozgun & Turan (2015), Gür (2015) and Aras & Sayın (2020).

Khairy & El-Shafay (2013) revealed that the highest protein of *Pterocladia capillacea* in the different seasons was  $23.72\pm0.03\%$  Mazlum, Yazıcı, Sayın, Habiboğlu & Ugur, (2021) and Aras & Sayın (2020) revealed that protein level of *Jania rubens* and *Ellisolandia elongata* from red macroalgae was  $5.99\pm0.773\%$  and  $6.05\pm0.03\%$ , respectively. Protein level of RMA(*Pterocladiella capillacea*) tested were higher than those of the levels determined by Mazlum et al. (2021) and Aras & Sayın (2020) but not Khairy & El-Shafay (2013) and Pakawan et al. (2015).

Burtin (2003) revealed that protein levels of GMA were generally between 10-30%. Firat, Öztürk, Taşkın & Kurt (2007) showed that protein value of *Caulerpa racemosa* were 12.94-20.18%. Manivannan, Thirumaran, Devi, Hemalatha & Anantharaman (2008) found that protein value of *Ulva intestinalis* was 16-17%. Manas et al. (2017) showed that protein value of *Caulerpa species* were 9.21-17.19%. Magdugo et al. (2020) showed that protein of *Caulerpa racemosa* was 19.9%.Protein level of *Ulva intestinalis* by Aras & Sayın (2020) determined as  $15.77\pm0.16\%$ . Protein value of *Ulva lactuca* belonging to GMA was determined as  $16.89\pm0.12\%$  by Mazlum et al. (2021). The protein values observed for GMA (*Chaetomorpha linum* and *Caulerpa prolifera*) in present study were lower than the values mentioned by Burtin, (2003); Fırat, et al. (2007); Manivannan et al. (2008); Pakawan et al. (2015); Manas et al. (2017); Magdugo et al. (2020); Aras & Sayın (2020) and Mazlum et al. (2021).

McDermid & Stuercke (2003) reported that lipid content of macroalgae was less than 4%. Polat & Ozogul (2008) showed that lipid levels of RMA and BMA were between 1.10-11.53%. Lipid level is generally low in macroalgae, between 1-5% (Peng et al., 2015).

Wahbeh (1997) revealed that the lipid content of the macroalgae *Padina pavonica* collected at the beach in Aqaba, Jordan was 4.4%. Gür (2015) and Sultana, Ambreen, & Tarıq (2012) showed that lipid levels of *Dictyota dichotoma* from BMA was 0.9-5.13% and 6.8%. Aras & Sayın (2020) revealed that lipid values of

*Dictyota dichotoma* and *Sargassum vulgare* were  $5.43\pm0.23\%$  and  $12.21\pm0.52\%$ , respectively. Ahmad, Sulaiman, Saimon, Yee, & Matanjun, (2012) stated that BMA species have higher lipid content than RMA and GMA species. Lipid levels in our study were between  $1.74\pm0.19\%$  (*Caulerpa prolifera*)- $5,83\pm0,68\%$  (*Ericaria amentacea*) levels stated by Peng et al. (2015). Uslu et al. (2021) determined that the lipid amounts of the macroalgae *Sargassum vulgare* and *Cystoseira compressa* were  $2.58\pm0.4\%$  and  $2.00\pm0.5\%$ , respectively. Lipid levels of *Sargassum vulgare* and *Cystoseira compressa* revealed by Uslu et al. (2021) were lower than present study. The results of our study were supported by Ahmad et al. (2012), Polat & Ozogul (2008), Peng et al. (2015), Gür (2015) but not Aras & Sayın (2020).

Mazlum et al. (2021) and Aras & Sayın (2020) revealed that lipid level of *Jania rubens* and *Ellisolandia elongata* from RMA were 0.39±0.103% and 0.43±0.09%, respectively. Lipid level of RMA(*Pterocladiella capillacea*) tested was higher than those of Mazlum et al. (2021) and Aras & Sayın (2020).

Khairy & El-Shafay (2013) showed that *Ulva lactuca* ( $4.09 \pm 0.2\%$ ) contained more lipids than *Jania rubens* and *Pterocladia capillacea*. Manas et al. (2017) showed that lipid value of Caulerpa species were 1.29-2.44%. Magdugo et al. (2020) determined that lipid value of *Caulerpa racemosa* was 4.5%. Mazlum et al. (2021) and Aras & Sayın (2020) revealed that lipid levels of *Ulva lactuca* and *Ulva intestinalis* were 1.08±0.33% and 1.04±0.37%, respectively. Lipid levels of GMA (*Chaetomorpha linum* and *Caulerpa prolifera*) tested were higher than those of Mazlum et al. (2021) and Aras & Sayın (2020). Results were supported by Polat & Ozogul (2008) and Peng et al (2015) and similiar to Khairy & El-Shafay (2013) except for *Caulerpa prolifera*.

According to the literatures, macroalgae have low lipid potential (Ratana- arporn & Chirapart, 2006). Chakraborty & Bhattacharya (2012) mentioned that lipid contents of macroalgae may vary depending on the type and amount of nutritive elements in the environment. Also, Peng et al. (2015) indicated that low lipid contents of macroalgae depends on light intensity, salinity and temperature conditions. Studies have shown that seasonal changes have caused changes in the biochemical compositions of macroalgae.

Tabarsa et al. (2012) revealed that the ash content of macroalgae varies between 8-40% of their dry weight. Polat & Ozogul (2013) determined that the ash levels varies between 2.28-51.63%. On the other hand, Liu (2017) showed that algae can contain as high as 70% dry matter ash in different locations.

Wahbeh (1997) indicated that the ash content of the macroalgae *Padina pavonica* collected at the beach in Aqaba, Jordan was 23.1%. Ozgun & Turan (2015) showed that the ash levels of 8 BMA gathered from Iskenderun Bay was varied from  $1.66\pm0.29\%$  to  $18.19\pm2.66\%$ . Uslu et al. (2021) determined that the ash amounts of the macroalgae *Sargassum vulgare* and *Cystoseira compressa* were  $27.05\pm0.5\%$  and  $21\pm0.1\%$ , respectively. Aras & Sayın (2020) found that the ash levels of BMA (*Dictyota dichotoma* and *Sargassum vulgare*) were  $27.34\pm0.72\%$  and  $14.79\pm0.19\%$ , respectively. BMA (*Sargassum vulgare* and *Ericaria amentacea*)) results were similar to the values determined by Wahbeh (1997), Ozgun & Turan (2015), and Aras & Sayın (2020) and Uslu et al. (2021) except for *Sargassum vulgare*.

Firat et al. (2007) revealed that ash value of *Caulerpa racemosa* were 8.02-19.50%. Manas et al. (2017) showed that ash value of *Caulerpa species* were 23.90-40.27%. Magdugo et al. (2020) revealed that ash value of *Caulerpa racemosa* was 29,4%. Aras & Sayin (2020) and Mazlum et al. (2021) revealed that the ash levels of GMA (*Ulva intstinalis*) and *Ulva lactuca*) were 27.49±0.43% and 26.47±0.20%. Aras & Sayin (2020) and Mazlum et al. (2021) determined that the ash levels of RMA (*Ellisolandia elongata* and *Janie rubens*) were 76.75±0.20% and 78.740±0.066%, respectively. Khairy & El-Shafay (2013) indicated that ash content of *Jania rubens* species were quite high (50.54%). Liu (2017) reveal that algae can contain as high as 70% dry matter ash in different locations. Results belonging to GMA (*Chaetomorpha linum* and *Caulerpa prolifera*) and RMA(*Pterocladiella capillacea*) were lower than those of Khairy & El-Shafay (2013), Liu (2017), Manas et al. (2017), Magdugo et al. (2020), Aras & Sayin (2020) and Mazlum et al. (2021). Factors such as geographical location and season might be important factors in change the ash content of macroalgae (Renaud and Luong-Van, 2006; Mohamed, Hashim & Rahman, 2012; Cabrita et al., 2016)

Aketa & Kawamura (2001) found that protein values of *Halophila ovalis* was 6,2%. Coria-Monter & Durán-Campos (2015) showed that the protein, lipid and ash values of three seagrass species (*Thalassia testudinum*, *Halodule wrightii*, and *Syringodium filiforme*) were 8.47%-8.10%-10.43%; 0.83%,-2.33%-2.13%; 38.77%-27.23%-23.43%, respectively. Protein level of Angiosperms/Seagrass (*Halophila stipulacea*) was similar to Aketa & Kawamura, (2001), Kannan, Arumugam & Anantharaman, (2010) and Coria-Monter & Durán-Campos (2015). However, lipid and ash values of *Halophila stipulacea* were higher and lower than those of seagrasses species reported by Kannan et al. (2010) and Kannan, Arumugam, Iyapparaj, Thangaradjou & Anantharaman (2013), respectively. Renaud & Luong-Van (2006) indicated that geographic location, environmental conditions, seasons, and sampling conditions changed the biochemical composition of seagrass.

## 4. Conclusion

In conclusion, the information of the biochemical compositions of five different macroalgae ((GMA (*Chaetomorpha linum* and *Caulerpa prolifera*), **RMA** (*Pterocladiella capillacea*), **BMA** (*Sargassum vulgare*. and *Ericaria amentacea*)) and Angiosperms/Seagrass (*Halophila stipulacea*) are important for the evaluation of potential sources for commercial and human consumption. Biochemical compositions of tested macroalgae and seagrass could make important contributions to feed formulations and fuctional foods in future.

## **Author Contributions**

Mehmet Naz: planned the study and statistical analyzes and evaulating the results and writing

Selin Sayın: planned the study and the writing of the manuscript

Zafer Çetin: biochemical analyses of macroalgae and the writing of the manuscript

Eyüp İlker Saygılı: biochemical analyses of macroalgae and the writing of the manuscript

Ergun Taşkın: collection of macroalgae and the writing of the manuscript

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## **Conflicts of Interest**

The authors declare no conflict of interest.

## References

- Ahmad, F., Sulaiman, M. R., Saimon. W., Yee, C. F., & Matanjun, P. (2012). Proximate compositions and total phenolic content of selected edible seaweed from Semporna, Sabah, Malaysia. *Borneo Science*, 31, 85-96. Retrieved from: https://jurcon.ums.edu.my/ojums/index.php/borneo-science/article/view/171
- Ak, İ. (2015). Sucul Ortamın Ekonomik Bitkileri Makro Algler. Dünya Gıda Dergisi, 12, 88-97. Retrieved from: https://www.academia.edu/20226751/Sucul\_Ortam%C4%B1n\_Ekonomik\_Bitkileri\_Makro\_Algler
- Aketa, K., & Kawamura, A. (2001). Digestive functions in Sirenians and others (Review). *The Bulletin of Faculty of Biosources*, 27, 85-103. Retrieved from: https://agris.fao.org/agris-search/search.do?re-cordID=JP2002004975
- AOAC (2005). Association of Official Analytical Chemist, Official Methods of Analysis. 18th Edition, AOAC International, Suite 500, 481 North Frederick Avenue, Gaithersburg, Maryland 20877-2417, USA. ISBN 0-935584-77-3
- Aras, A., & Sayın, S. (2020). Geleceğin Fonksiyonel Ürünleri için Bazı Denizel Makroalglerin Potansiyellerinin Belirlenmesi. *MedFAR*, 3(1), 22-35. Retrieved from: https://dergipark.org.tr/tr/pub/medfar/issue/52185/640946

- Benjama, O., & Masniyom, P. (2011). Nutritional composition and physicochemical properties of two green seaweeds (*Ulva pertusa* and *Ulva intestinalis*) from the Pattani Bay in Southern Thailand. Songklanakarin. J. Sci. Technol., 33, 575–583. Retrieved from: https://agris.fao.org/agris-search/search.do?recordID=AV2012081280
- Bligh, E.G., & Dyer, W.J. (1959). A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.*, 37, 911–917. Retrieved from: https://cdnsciencepub.com/doi/10.1139/o59-099
- Burtin, P. (2003). Nutritional value of seaweeds. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 2: 498-503. Retrieved from: https://www.semanticscholar.org/paper/Nutritional-value-of-seaweeds-Burtin/32f330e9136c9a9ffc794217f50d4814c7b59326
- Cabrita, A.R.J., Maia, M.R.G., Oliveira, H.M., Sousa-Pinto, I., Almeida, A.A., Pinto, E., & Fonseca, A.J.M. (2016). Tracing seaweeds as mineral sources for farm-animals. *The Journal of Applied Phycology*, 28, 3135–3150. doi: https://doi.org/10.1007/s10811-016-0839-y
- Chakraborty, S., & Bhattacharya, T. (2012). Nutrient composition of marine benthic algae found in the gulf of Kutch Coastline, Gujarat, India. *Journal of Algal Biomass Utilization*, 3 (1), 32 – 38. Retrieved from: https://www.semanticscholar.org/paper/Nutrient-composition-of-marine-benthic-algae-found-Chakraborty-Bhattacharya/868e550a4149a1456822b56c48096a53ae14e437
- Coria-Monter, E., & Durán-Campos E. (2015). Proximal analysis of seagrass species from Laguna de Términos, Mexico. *Hidrobiológica*, 25 (2), 249-255. Retrieved from: https://www.semanticscholar.org/pa-per/Proximal-analysis-of-seagrass-species-from-Laguna-Coria-Monter-Dur%C3%A1n-Campos/18fa4c4d863e1c5bce006b79334a06daf70a4eb2
- Cormaci, M., Furnari, G., & Alongi, G. (2014). Flora marina bentonica del Mediterraneo: Chlorophyta. Boll. Accad. Gioenia Sci. Nat. Catania 47, 11–436. Retrieved from: https://www.researchgate.net/publication/311886135\_Flora\_marina\_bentonica\_del\_Mediterraneo\_Chlorophyta
- Dawczynski, C., Schubert, R., & Jahreis, G. (2007). Amino acids, fatty acids, and dietary fibre in edible seaweed products. *Food Chemistry*, 103(3), 891–899. doi: https://doi.org/10.1016/j.foodchem.2006.09.041
- Fırat, C., Öztürk, M., Taşkın, E. & Kurt, O. (2007). Caulerpa racemosa (Forsskal) J. Agardh'nın (Chlorophyceae=Yeşil Algler) Biyokimyasal İçeriği. Ege Journal of Fisheries and Aquatic Sciences, 24 (1), 89-91. Retrieved from: https://app.trdizin.gov.tr/makale/TnpjME5EQXc/caulerpa-racemosa-forsskal-jagardh-nin-chlorophyceae-yesil-algler-biyokimyasal-icerigi
- Fourqurean, J.W., Duarte, C.M., Kennedy, H., Marba, N., Holmer, M., Mateo, M.A., Apostolaki, E.T., Kendrick, G.A., Krause-Jensen, D., McGlathery, K.J., & Serrano, O. (2012).Seagrass ecosystems as a globally significant carbon stock. *Nat. Geosci.*, 5, 505–509. doi: https://doi.org/10.1038/ngeo1477
- Gür, İ. (2015). Investigation on the seasonal variations of pigments, antioxidants and nutritional composition in some macroalgae species distrubuted in the Iskenderun Bay. *Cukurova University Institute of Natural* and Applied Sciences, MSc Thesis, 71 p. Retrieved from: https://tez.yok.gov.tr/UlusalTezMerkezi/tez-SorguSonucYeni.jsp
- Hemminga, M.A., & Duarte, C.M. (2000). Seagrass Ecology. Cambridge University Press, Cambridge, UK, 298 doi: https://doi.org/10.1017/CBO9780511525551
- Kannan R., Arumugam, R., & Anantharaman, P. (2010). Antibacterial potential of three seagrasses against human pathogens. Asian Pac. J. Trop. Med., 3, 890-893. doi: https://doi.org/10.1016/S1995-7645(10)60214-3
- Kannan, R., Arumugam, R., Iyapparaj, P., Thangaradjou, T., & Anantharaman, P. (2013). In vitro antibacterial, cytotoxicity and haemolytic activities and phytochemical analysis of seagrasses from the Gulf of Mannar, South India. *Food Chem.*, 136, 1484-1489. doi: 10.1016/j.foodchem.2012.09.006
- Khairy H. M., and El-Shafay S. M. (2013). Seasonal variations in the biochemical composition of some common seaweed species from the coast of Abu Qir Bay, Alexandria, Egypt. *Oceanologia*, 55(2): 435–452. doi: https://doi.org/10.5697/oc.55-2.435

- Lipkin, Y., (1975). *Halophila stipulacea*, a review of a successful immigration. *Aquat. Bot.*, 1, 203–215. doi: https://doi.org/10.1016/0304-3770(75)90023-6
- Liu, K. (2017). Characterization of ash in algae and other materials by determination of wetacid indigestible ash and microscopic examination. *Algal Research*, 25, 307-321. doi: https://doi.org/10.1016/j.al-gal.2017.04.014
- Magdugo, R. P., Terme, N., Lang, M., Pliego-Cortés, H., Marty, C., Hurtado, A.Q., Bedoux, G., & Bourgougnon, N. (2020). An Analysis of the Nutritional and Health Values of *Caulerpa racemosa* (Forsskål) and *Ulva fasciata* (Delile)—Two Chlorophyta Collected from the Philippines. *Molecules*, 25 (12), 2901. doi: 10.3390/molecules25122901
- Malm, T. (2006). Reproduction and recruitment of the seagrass *Halophila stipulacea*. Aquat. Bot., 85, 345–349. doi: https://doi.org/10.1016/j.aquabot.2006.05.008
- Manas, H. M., Deshmukhe, G., Venkateshwarlu, G., Jaiswar, A. K., Divipala, I., Siddaiah G. M. (2017).Nutritional profile of some important Caulerpa J.V. Lamouroux species along Maharashtra and Gujarat coast, India. *Journal of Fisheries and Life Sciences*, 2(1), 30-34. Retrieved from: https://www.semanticscholar.org/paper/Nutritional-profile-of-some-important-Caulerpa-J.V.-Manas-Deshmukhe/9e7320989c69560cff4e83b700ba6de686b14aa5
- Manivannan, K., Thirumaran, G., Devi, G.K., Hemalatha, A., & Anantharaman, P. (2008). Biochemical composition of seaweeds from mandapam coastal regions along southeast coast of India. *American-Eurasian Journal of Botany.*, 1 (2), 32-37. Retrieved from: https://www.semanticscholar.org/paper/Biochemical-Composition-of-Seaweeds-from-Mandapam-Manivannan-Thiruma-ran/69cce43a111aab691880d7ed710aa3cd48c9c4bd
- Mazlum, Y., Yazıcı, M., Sayın, S., Habiboğlu, O., & Ugur, S. (2021). Effects of two different macroalgae (*Ulva lactuca* and *Jania rubens*) species on growth and survival of juvenile red swamp crayfish (*Procambarus clarkii*) as feed additive. *Mar. Sci. Tech. Bull.*, 10(2), 154-162. Retrieved from: https://dergipark.org.tr/tr/download/article-file/1378380
- Mcdermid, K. J., & Stuercke, B., (2003). Nutritional composition of edible Hawaiian seaweeds. Journal of Applied Phycology., 15, 513–524.10.1023/B:JAPH.0000004345.31686.7f
- Mohamed, S., Hashim, S.N., & Rahman, H.A. (2012). Seaweeds: a sustainable functional food for complementary and alternative therapy. *Trends Food Sci. Tech.*, 23, 83-96. doi: https://doi.org/10.1016/j.tifs.2011.09.001
- Özgün, S. & Turan, F. (2015). Biochemical composition of some brown algae from Iskenderun Bay, The Northeastern Mediterranean coast of Turkey. *Journal of the Black Sea / Mediterranean Environment*, 2, 125-134.https://blackmeditjournal.org/wp-content/uploads/Funda\_Turan.pdf
- Pakawan, S., Suriyan, T., Kriengkrai, S., & Jintana, S. (2015). Growth and Nutrients Analysis in Marine Macroalgae. *Kasetsart J. (Nat. Sci.)*, 49, 211-218. Retrieved from: https://li01.tci-thaijo.org/index.php/anres/article/view/243563
- Peng, Y., Hu, J., Yang, B., Lin, X. P., Zhou, X. F., Yang, X.W., & Liu, Y. (2015). Chemical composition of seaweeds. *In: Seaweed Sustainability. Academic Press*, 79-124. doi: https://doi.org/10.1016/B978-0-12-418697-2.00005-2
- Polat, S., & Ozogul, Y. (2008). Biochemical composition of some red and brown macro algae from the Northeastern Mediterranean Sea. *International Journal of Food Sciences and Nutrition*, 59(7-8), 566-572. doi: https://doi.org/10.1080/09637480701446524
- Polat, S., & Özoğul, Y. (2013). Seasonal proximate and fatty acid variations of some seaweeds from the Northeastern Mediterranean coast. *Oceanologia*, 55 (2), 375–391. doi:10.5697/OC.55-2.375
- Pradana G B., Prabowo K B, Hastuti R P., Djaeni M., Prasetyaningrum A., 2019. Seaweed Drying Process Using Tray Dryer with Dehumidified Air System to Increase Efficiency of Energy and QualityProduct IOP Conf. Series: Earth and Environmental Science 292, 012070. Retrieved from: https://iopscience.iop.org/article/10.1088/1755-1315/292/1/012070
- Renaud, S.M., & Luong-Van, J.T. (2006). Seasonal variation in the chemical composition of tropical Australian marine macroalgae. J. Appl. Phycol., 18,381-387. doi: https://doi.org/10.1007/s10811-006-9034-x

- Ratana-arporn, P., & Chirapart, A. (2006). Nutritional evaluation of tropical green seaweeds *Caulerpa lentillifera* and *Ulva reticulata. Kasetsart J. (Nat. Sci.)*, 40,75–83. Retrieved from: https://www.cabi.org/ISC/abstract/20073167377
- Rodríguez-Prieto, C., Ballesteros, E., Boisset, F. & Afonso-Carrillo, J. (2013). Guía de las macroalgas y fanerógamas marinas del Mediterráneo occidental. pp. 656. Barcelona: Ediciones Omega, Retrieved from: https://www.amazon.es/Macroalgas-Faner%C3%B3gamas-Mediterr%C3%A1neo-Occidental-NATU-RALISTA-PECES/dp/8428215928
- Sultana, V., Ambreen, H.K., & Tarıq, A. (2012). Evaluation of biochemical component and antimicrobial activity of some seaweeds occuring at Karachi coast. *Pakistan Journal of Botany*, 44(5), 1799-1803. Retrieved from: https://agris.fao.org/agris-search/search.do?recordID=PK2017000363
- Tabarsa, M., Rezaei, M., Ramezanpour, Z., Robert Waaland, J., & Rabiei, R. (2012). Fatty acids, Amino acids, Mineral contents and Proximate composition of Some Brown Seaweeds. *Journal of Phycology*, 48(2), 285-292. doi: 10.1111/j.1529-8817.2012.01122.x
- Taşkın, E., & Öztürk, M. (2013). Türkiye Deniz Algleri. I. Phaeophyceae., p. 1-229, Manisa, Türkiye. Retrieved from: https://www.researchgate.net/publication/274196118\_Turkiye\_Deniz\_Algleri\_I\_Phaeophyceae
- Uslu, L., Sayın, S., Naz, M., Taskın, E., Soyler, O., Saygılı İ., Çetin, Z., Dinler, ZM., & Işık O. (2021). Proximate Analysis and Fatty Acid Profile of Some Brown Macroalgae Collected from the Northeastern Mediterranean Coast. *Fresenius Environmental Bultein*, 30(7), 9433-9437. Retrieved from: https://www.prt-parlar.de/download\_feb\_2021/
- Wahbeh, M.I. (1997). Amino acid and fatty acid profiles of four species of macroalgae from Aqaba and their suitability for use in fish diets. *Aquaculture*, 159(1-2), 101-109. Retrieved from: https://doi.org/10.1016/S0044-8486(97)00183-X
- Wells, M.L., Potin, P., Craigie, J.S., Raven, J.A., Merchant, S. S., Helliwell, K. E., & Brawley, S. H. (2017). Algae as nutritional and functional food sources: revisiting our understanding. *Journal of Applied Phycology*, 29, 949–982. doi: 10.1007/s10811-016-0974-5